

UNCLASSIFIED

AD NUMBER
AD860429
NEW LIMITATION CHANGE
TO Approved for public release, distribution unlimited
FROM Distribution authorized to U.S. Gov't. agencies and their contractors; Administrative/Operational Use; SEP 1969. Other requests shall be referred to Department of the Army, Fort Detrick, Attn: Technical Release Branch/TID, Frederick, MD 21701.
AUTHORITY
BDRL D/A ltr, 29 Sep 1971

THIS PAGE IS UNCLASSIFIED

AD

TECHNICAL MANUSCRIPT 559

CANDIDIASIS IN SIMIANS

Steven E. Wikse

James G. Fox

Robert M. Kovatch

STATEMENT #2 UNCLASSIFIED

This document is subject to special export controls and each transmittal to foreign governments or foreign nationals may be made only with prior approval of Dept. of Army, Fort Detrick, ATTN: Technical Release Branch/TID, Frederick, Maryland 21701

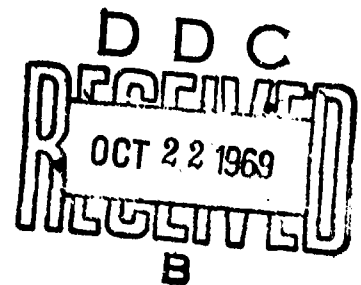
SEPTEMBER 1969

DEPARTMENT OF THE ARMY

Fort Detrick

Frederick, Maryland

Reproduced by the
CLEARINGHOUSE
for Federal Scientific & Technical
Information Springfield Va. 22151



Reproduction of this publication in whole or in part is prohibited except with permission of the Commanding Officer, Fort Detrick, ATTN: Technical Releases Branch, Technical Information Division, Fort Detrick, Frederick, Maryland, 21701. However, DDC is authorized to reproduce the publication for United States Government purposes.

DDC AVAILABILITY NOTICES

Qualified requesters may obtain copies of this publication from DDC.

Foreign announcement and dissemination of this publication by DDC is not authorized.

Release or announcement to the public is not authorized.

DISPOSITION INSTRUCTIONS

Destroy this publication when it is no longer needed. Do not return it to the originator.

The findings in this publication are not to be construed as an official Department of the Army position, unless so designated by other authorized documents.

TESTI	WHITE SECTION	<input type="checkbox"/>
DOC	DIFF SECTION	<input checked="" type="checkbox"/>
U. ANNOUNCED		<input type="checkbox"/>
JUSTIFICATION		
BY		
DISTRIBUTION/AVAILABILITY CODES		
DIST.	AVAIL.	AND OR SPECIAL
2		

DEPARTMENT OF THE ARMY
Fort Detrick
Frederick, Maryland 21701

TECHNICAL MANUSCRIPT 559

CANDIDIASIS IN SIMIANS

Steven E. Wikse

James G. Fox

Robert M. Kovatch

Pathology Division
MEDICAL SCIENCES LABORATORIES
and
Animal Farm Division
AEROBIOLOGY & EVALUATION LABORATORIES

Project 1B562602AD01

September 1969

In conducting the research described in this report, the investigators adhered to the "Guide for Laboratory Animal Facilities and Care," as promulgated by the Committee on the Guide for Laboratory Animal Facilities and Care of the Institute of Laboratory Animal Resources, National Academy of Sciences-National Research Council.

ABSTRACT

Candidiasis was diagnosed in six monkeys over a 10-month period. Most cases had been on antibiotic therapy for enterocolitis. Fungal invasion was seen in epithelium of the tongue, oral cavity, esophagus, and colon, and in hard keratin of the nails. Gross lesions of the anterior alimentary tract were either white patches or ulcers of the mucosa. Lesions of the colon consisted of a thick pseudomembrane that contained numerous Candida. The nails exhibited typical Candida onychomycosis. C. albicans was isolated from the two cases that were cultured. Tissue invasion by Candida blastospores and hyphae was histologically demonstrated in all cases.

I. INTRODUCTION*

Fungi of the genus Candida are common saprophytes of the skin, alimentary tract, and reproductive tract of monkeys. Candida spp. were isolated from 8 to 100% of healthy monkeys cultured in 10 surveys¹⁻¹⁰ and C. albicans was the species most frequently recovered. The common sites of isolation were the oral cavity, rectum, vagina, and skin.

As a saprophyte, Candida is found in keratin of the stratum corneum of epithelium and in digestive tract contents. Occasionally it has been isolated from soil or vegetable sources and is considered an obligate saprophyte because it does not propagate outside the body.¹¹ Numerous debilitating conditions enable this facultative pathogen to produce serious and even fatal disease in primates.

We have found reports in the literature of six cases of candidiasis in subhuman primates. Candida albicans has been isolated from tongue lesions by Thiry¹² in a vervet monkey (Ceracopithecus patas) experimentally infected with Trypanosoma cruzi and by Fiennes¹³ in a woolly monkey (Lagothrix sp.) and a gorilla (Gorilla gorilla beringei). Fiennes also recovered C. albicans from the tongue, mouth, lungs, liver, and intestine of a capuchin (Cebus sp.) that died of systemic candidiasis. Saëz¹⁴ cultured C. albicans from a barbary ape (Macaca sylvana) with a lung condition. Kerber, Reese, and Van Natta¹⁵ isolated C. albicans from the penis and hand of a rhesus monkey (Macaca mulatta) with balanitis, paronychia, and onychia that responded to antifungal therapy.

We feel that the frequency of clinical candidiasis in simians is greater than the literature indicates. The present paper describes the clinical signs, gross and microscopic lesions, and diagnostic mycology of spontaneous candidiasis in six monkeys.

* This report should not be used as a literature citation in material to be published in the open literature. Readers interested in referencing the information contained herein should contact the senior author to ascertain when and where it may appear in citable form.

II. MATERIALS AND METHODS

Cases 1 to 5 are rhesus monkeys (*M. mulatta*) that became ill during the 3-month conditioning period at the Fort Detrick Animal Farm. Case 6 is a capuchin (*Cebus albifrons*) that died at the quarantine facility of a local pet shop. The Fort Detrick monkeys were maintained on a preventive disease regimen consisting of tuberculin testing and intramuscular injection of benzathine penicillin G upon arrival, prophylactic levels of oxytetracycline* in the feed** for 2 weeks after arrival, and administration of thiabendazole 6 and 9 weeks following arrival. Standard diarrhea therapy consisted of intramuscular vitamin B complex and antibiotics combined with an oral preparation of furazolidone, kaolin pectate, electrolytes,*** and 10% dextrose administered via stomach tube. Chloramphenicol, kanamycin, and ampicillin were the commonly administered antibiotics. Duration of treatment varied from 2 to 6 consecutive days. Bacterial culture, antibiotic sensitivity tests, egg flotation, and direct smears for parasites were conducted on fecal samples of nonresponsive cases.

Monkeys with ulcers or elevated white plaques of the tongue or oral cavity were considered candidiasis suspects. Two cases were clinically diagnosed by direct smears of lesions in 10% NaOH stained with methylene blue and cultured on Sabouraud's agar. *C. albicans* was identified by sugar fermentation tests¹⁶ in case 5 and by production of chlamydospores on chlamydospore agar**** in case 6.

To determine virulence, rabbits were injected with the isolate from case 6. Serial tenfold dilutions were made from a saline suspension of a 48-hour culture grown on Sabouraud's agar. Three-pound rabbits were injected intracardially with 0.5 ml of the 10^{-1} , 10^{-2} , 10^{-3} , and 10^{-5} dilutions. To estimate the number of organisms inoculated, 0.1 ml of each dilution was streaked on plates of Sabouraud's agar.

All monkeys were necropsied and tissue sections were routinely stained with hematoxylin and eosin; Gomori methenamine silver or Gridley's fungus stains were used to vividly color fungi.¹⁷

* Terramix A-B-D-25[®], Chas. Pfizer & Co., Inc., New York, New York.

** Purina Monkey Chow, Ralston Purina Co., St. Louis, Mo.

*** Vetrad-O[®], Don Hall Laboratories, Norwich, New York.

**** Chlamydospore agar, Baltimore Biological Laboratories, Baltimore, Md.

III. RESULTS

Candidiasis was diagnosed in the six monkeys over a period of 10 months. Predisposing conditions were present in five cases. Cases 1 to 4 had dysentery and had undergone several days' antibiotic and symptomatic therapy prior to death. Cases 1 and 3 had been successfully treated for dysentery several weeks before the final illness. Case 6 had a systemic viral infection microscopically characterized by intranuclear and cytoplasmic, eosinophilic inclusion bodies in reticuloendothelial cells of the lymphoid system. Case 5 was dyspneic and C. albicans was isolated from pharyngeal ulcers. This monkey was sacrificed to determine the relationship of the fungus to the lesions.

Clinical signs of candidiasis varied with the location of lesions. Fungal invasion was seen in epithelium of the tongue, oral cavity, esophagus, and colon, and in hard keratin of the nails. Ulcers or white, slightly elevated, rough-surfaced plaques of the tongue or oral cavity were seen on physical examination in cases 2, 5, and 6. Plaques varied in size from 0.3 to 1 cm in diameter in case 5 to a white covering of the entire dorsum of the tongue in case 6. The lesions sometimes were difficult to see grossly, as in case 2, which had mild superficial sloughing of small areas of the posterior dorsum of the tongue. Open-mouth breathing occurred in case 5, which had ulcers 0.5 to 1 cm in diameter on a swollen soft palate. Case 2 had a severe ulcerative esophagitis that caused dysphagia. Upon necropsy each of the above cases had oral lesions and white longitudinal streaks 1 to 2 mm wide in the esophageal mucosa. Microscopically, the white plaques consisted of Candida and bacteria multiplying in a greatly thickened stratum corneum (Fig. 1). Oval blastospores were located near the surface and thin septate hyphae penetrated the stratum spinosum (Fig. 2). Neutrophils were located in pockets of the stratum corneum and in clefts between it and the stratum spinosum. There were hyperplastic rete pegs and a mild infiltration of mononuclear cells in the dermal papillae in involved areas (Fig. 3). Cases 1 and 3 had microscopic candidiasis of the tongue. Invasion of the lamina propria by hyphae was seen in the ulcerated esophagus of case 2 (Fig. 4-6). Cellular debris and myriad blastospores partially occluded its lumen.

The colon was involved in cases 2 and 4. The lumen of the terminal descending colon and rectum of case 4 was partially occluded by a thick layer of firm pink material that adhered to the mucosa. Microscopically this material was a pseudomembrane consisting of degenerating epithelial cells, leukocytes, bacteria, and large numbers of Candida blastospores and hyphae lodged in an acidophilic proteinaceous matrix (Fig. 7 and 8). The underlying mucosa was acutely inflamed. Massive Candida colonization was seen in the lumen of the inflamed colon of case 2.



FIGURE 1. Dorsolateral Portion of Tongue with Greatly Thickened Stratum Corneum. Case 6; H & E, 42X.

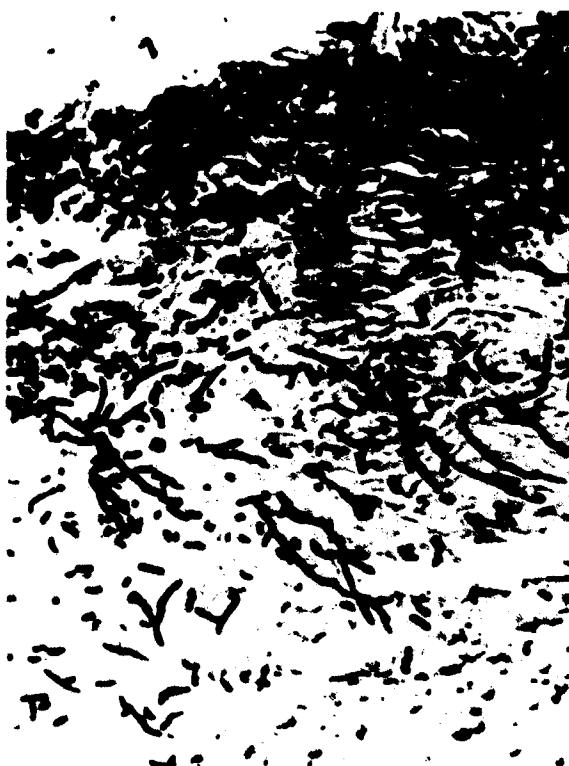


FIGURE 2. Blastospores Near the Surface and Hyphae Penetrating Through the Thickened Stratum Corneum into the Stratum Spinosum of the Tongue. Case 6; GMS, 130X.

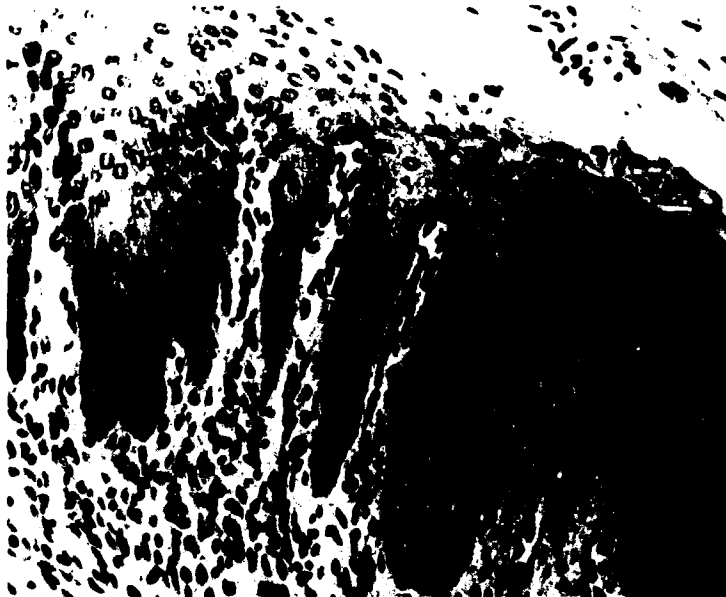


FIGURE 3. Hyperplastic Rete Pegs and Infiltration of Mononuclear Cells in Dermal Papillae of the Epithelium of the Tongue. Case 6; H & E, 420X.

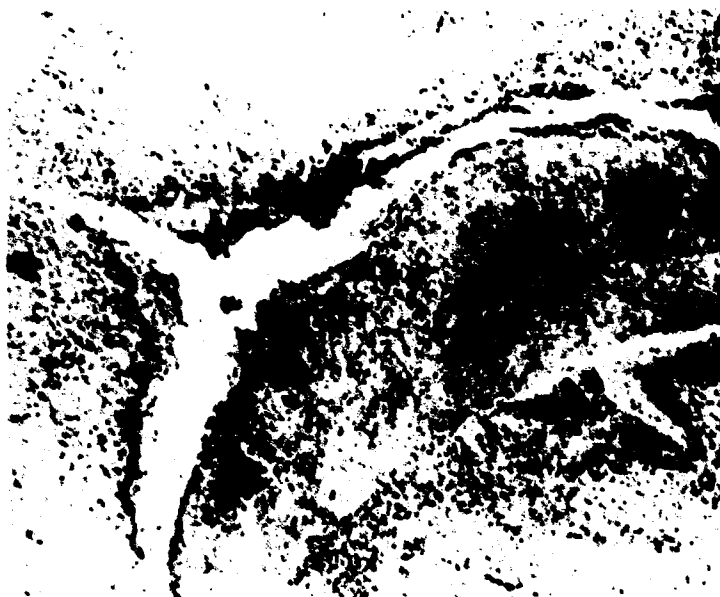


FIGURE 4. Ulcerated Esophagus with Destruction of Most of the Mucosal Epithelium. The lumen is partially occluded by cellular debris and Candida. Case 2; H & E, 42X.

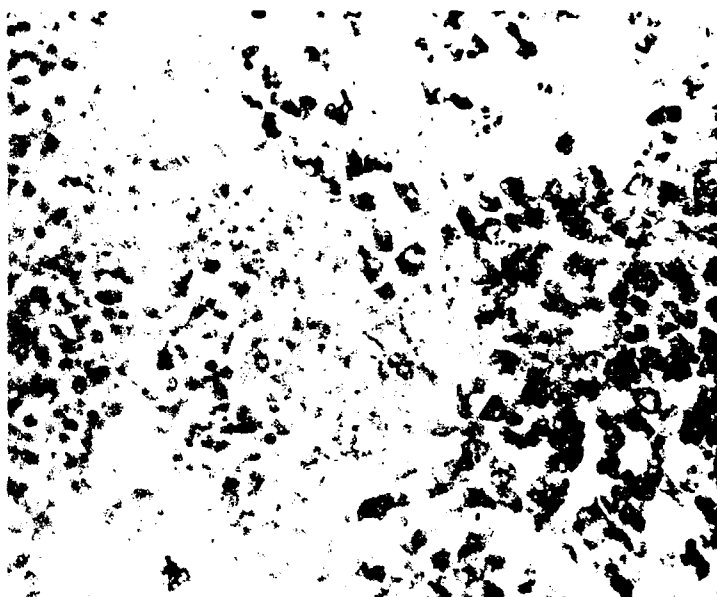


FIGURE 5. Numerous Candida Blastospores Adjacent to Degenerating Inflammatory Cells within the Esophagus. Case 2; H & E, 420X.

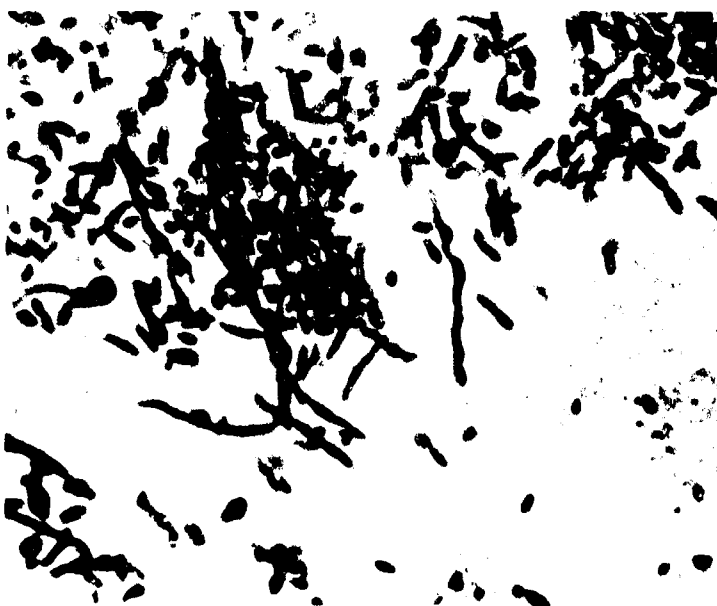


FIGURE 6. Hyphae of Candida Penetrating the Lamina Propria of the Ulcerated Esophagus. Case 2; GMS, 420X.

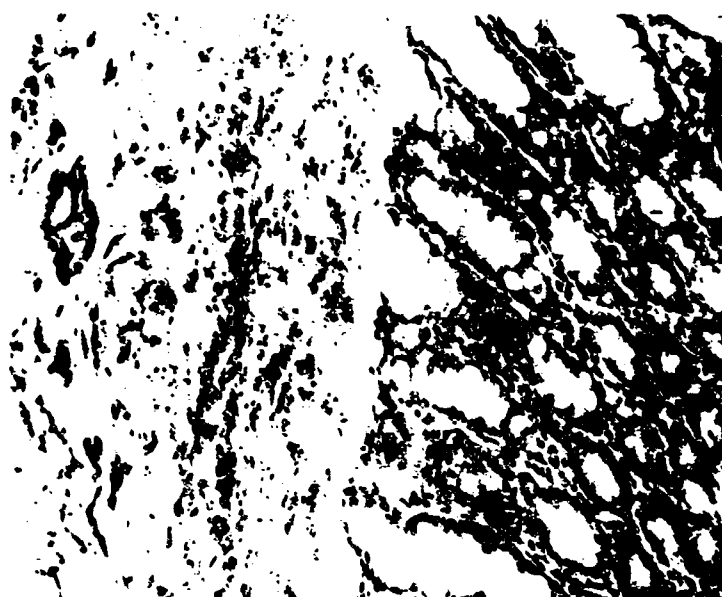


FIGURE 7. Pseudomembrane Adherent to Inflamed Colon. The proteinaceous material within dilated crypts of Lieberkühn is identical to the ground substance of the pseudomembrane. Case 4; H & E, 130X.

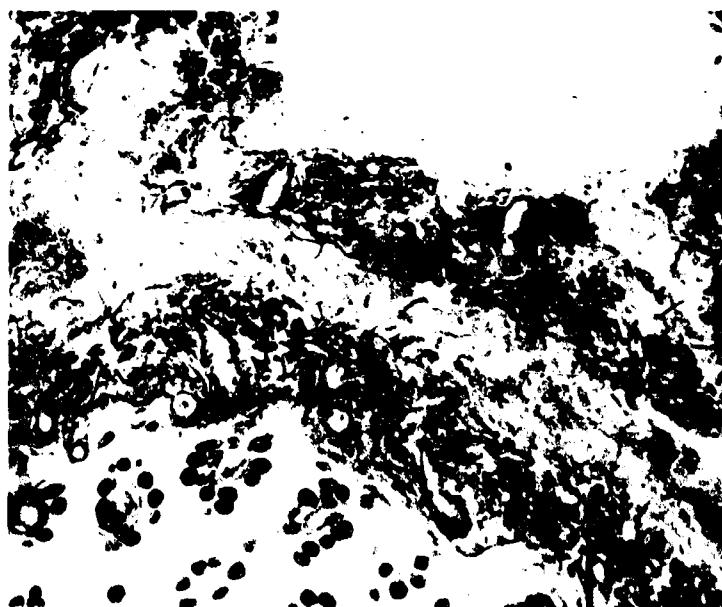


FIGURE 8. Blastospores and Hyphae in Pseudomembrane of the Colon. Case 4; GMS, 130X.

Candida onychomycosis occurred in case 1. The fingernails and toenails were eroded, shortened, and deformed. This monkey also had circular shallow ulcers on the fingertips and toes and a focally inflamed prepuce. Microscopic examination of the deformed nails revealed a disruption of the layers of keratin by Candida blastospores and hyphae (Fig. 9 and 10). The digital ulcers and posthitis were primarily bacterial inflammations with secondary surface growth of Candida.

Candidiasis was clinically diagnosed in cases 5 and 6. Filamentous hyphae and budding yeasts were seen in direct smears of lesions stained with methylene blue. Culture on Sabouraud's agar at 25 C produced smooth, raised, white colonies with a yeast-like odor within 36 hours. The isolates were identified as C. albicans in case 5 by the production of acid and gas in selected sugars¹⁵ and in case 6 by the production of chlamydospores (Fig. 11). Approximately 375,000 organisms of this isolate* inoculated intracardially into a rabbit produced death in 5 days, with disseminated abscesses 1 to 2 mm in diameter. The organism was recovered by culture from the abscesses and microscopically demonstrated in tissue section.

IV. DISCUSSION

These cases illustrate several general features of Candida infections. This opportunistic fungus is capable of living as a saprophyte or pathogen. Candidiasis, with serious, and sometimes fatal, manifestations has occurred with many conditions that result in an impairment of the body's defense mechanisms. Neoplastic and infectious diseases, immunologic defects, and conditions requiring antibiotic, corticosteroid, antimetabolite, or radiation therapy are included in this category.¹⁸ The examples presented are cases of infectious disease treated with antibiotics, with subsequent invasion by Candida. Recently there have been excellent discussions on the interrelationships of Candida, bacteria, and antibiotics.^{18,19}

The role of Candida as a secondary invader should not diminish our concept of its significance. We feel that its clinical effects can be more severe than those of the suspected underlying process. The ulcerative esophagitis of case 2 contributed to its fatal outcome. Esophageal candidiasis should be considered in dysphagic monkeys, especially if oral candidiasis lesions are present. Candida produces a potent endotoxin-like substance that has produced fatal shock when injected intravenously in mice.^{20,21} The extent of local damage or adverse systemic effects by this toxin in instances of massive intestinal colonization as seen in cases 2 and 5 is not known.

* Identification of culture from case 6 was confirmed by National Animal Disease Laboratory, Ames, Iowa.

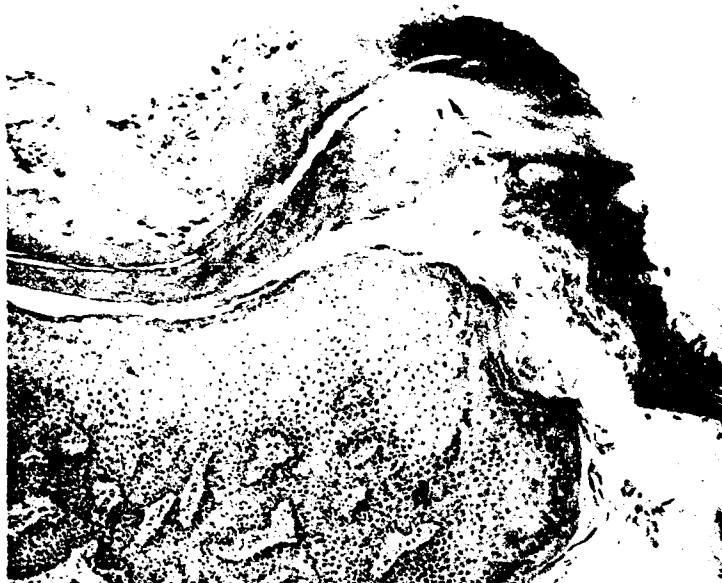


FIGURE 9. Shortened, Deformed Fingernail. Case 1; H & E, 42X.



FIGURE 10. Blastospores and Hyphae of Candida within the Keratin of the Nail. Case 1, Gridley's fungus stain, 420X.

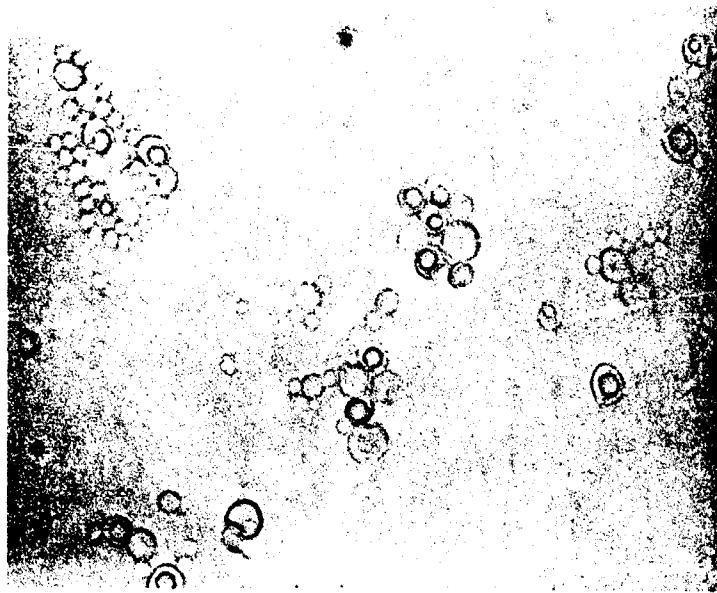


FIGURE 11. Thick-Walled Chlamydospores Characteristic of *C. albicans* Grown on Chlamydospore Agar. Case 6; slide culture, 130X.

Differential diagnosis of Candida tongue lesions includes ulcers associated with herpes virus infections²² and monkey pox.²³ The common saprophytic occurrence of Candida throughout the body limits the diagnostic value of cultures. Cultures are considered pathognomonic only when they originate from a tissue or fluid, such as blood, that does not normally harbor Candida.²⁴ It is thought that in vivo the yeast form of Candida is saprophytic and the hyphal form is invasive.²⁵ The demonstration of hyphae in fecal smears indicates intestinal candidiasis.²⁶ In biopsy or necropsy tissue sections, blastospores and hyphae can be identified satisfactorily as members of the genus Candida.²⁷ Species identification requires culture and differential tests.

Superficial candidiasis can be treated successfully if antifungal therapy is combined with correction of predisposing factors. Skin and vaginal infections respond well to topical nystatin or chlordantoin.²⁸ Nail infections are generally refractory to treatment, although nystatin is sometimes effective.^{15,16} Oral nystatin is very effective for digestive tract candidiasis.²⁶ The drug of choice for systemic candidiasis, which has a poor prognosis, is intravenous amphotericin B.²⁸

A correlation between positive Candida and tuberculin skin tests in guinea pigs harboring Candida was found by Vogel et al.²⁹ Matsufuji, Fukunda, and Nakamura³⁰ demonstrated in vivo that a common antigen is shared by C. albicans and Mycobacterium tuberculosis. This raises the possibility that C. albicans may be the cause of false positive tuberculin reactions in monkeys.

LITERATURE CITED

1. Al-Doory, Y. 1966. The mycoflora of the intestinal contents of the vervet monkey. *Mycologia* 58:659-662.
2. Al-Doory, Y. 1967. Microbiological parameters of the baboon (Papio sp.): *Mycology*, p. 731-739. In H. Vagtborg (ed.) *The baboon in medical research*, Vol. 2. University of Texas Press, Austin, Texas.
3. Al-Doory, Y. 1967. The mycoflora of the subhuman primates:
I. The flora of the oral cavity of the baboon in captivity. *Mycopathol. Mycol. Appl.* 31:43-48.
4. Al-Doory, Y. 1967. The mycoflora of the subhuman primates:
II. The flora of the rectum and vagina of the baboon in captivity. *Mycopathol. Mycol. Appl.* 31:332-336.
5. Al-Doory, Y. 1967. The mycoflora of the subhuman primates:
III. The flora of the skin of the baboon, vervet, and gelada in captivity. *Folia Primatol.* 7:292-298.
6. Cantanei, A. 1925. Presence d'un Monilia sur la langue de singes d'Algerie. *Compt. Rend. Soc. Biol.* 93:92-94.
7. Kohler, G. 1964. Intestinal yeast flora in captive zebras and monkeys. *Berlin. Muench. Tieraerztk. Wochensch.* 77:462-464.
8. Parle, J.N. 1957. Yeasts isolated from the mammalian alimentary tract. *J. Gen. Microbiol.* 17:363-367.
9. Van Uden, N. 1960-1961. The occurrence of Candida and other yeasts in the intestinal tracts of animals. *Ann. N.Y. Acad. Sci.* 89:59-68.
10. Wawrzkieicz, K.; Pietrzyk, J. 1965. Fungi of the genus Candida occurring in the oral cavity and vagina in monkeys. *Laboratory Animals (Polish)* 4:85-101.
11. Maddy, K.T. 1967. Epidemiology and ecology of deep mycoses of man and animals. *Arch. Dermatol.* 96:409-417.
12. Thiry, G. 1913. Muquet spontané chez le singe. *Arch. Parasitol.* 16:168-176.
13. Fiennes, R. 1967. Zoonoses of primates, p. 86. Weidenfeld and Nicolson, London.
14. Saëz, H. 1960. Champignons isolés du poumon de quelques mammifères sauvages morts en captivité. *Parassitologia* 2:353-358.

15. Kerber, W.T.; Reese, W.H.; Van Natta, J. 1968. Balanitis, paronychia, and onychia in a rhesus monkey. *Lab. Animal Care* 18:506-507.
16. Ajello, L.; George, L.K.; Kaplan, W.; Kaufman, L. 1966. Laboratory manual for medical mycology. U.S. Department of Health, Education and Welfare, Public Health Service, Communicable Disease Center, Atlanta, Georgia.
17. Armed Forces Institute of Pathology. 1968. Manual of histologic staining methods, 3rd ed. H.T. Luna (ed.) McGraw-Hill Book Company, New York.
18. Seelig, M.S. 1966. The role of antibiotics in the pathogenesis of Candida infections. *Amer. J. Med.* 40:887-917.
19. Seelig, M.S. 1966. Mechanisms by which antibiotics increase the incidence and severity of candidiasis and alter the immunological defenses. *Bacteriol. Rev.* 30:442-459.
20. Mourad, S.; Friedman, L. 1961. Pathogenicity of Candida. *J. Bacteriol.* 81:550-556.
21. Roth, F.J.; Murphy, W.H. 1957. Lethality of cell-free extract of Candida albicans for chlortetracycline-treated mice. *Proc. Soc. Exp. Biol. Med.* 94:530-532.
22. Hunt, R.D.; Melendez, L.V. 1969. Herpes virus infections of non-human primates: A review. *Lab. Animal Care* 19:221-234.
23. Sauer, R.M.; Prier, J.E.; Buchanan, R.S.; Creamer, A.A.; Fegley, H.C. 1960. Studies on a pox disease of monkeys: I. Pathology. *Amer. J. Vet. Res.* 21:377-380.
24. Kozinn, P.J.; Taschdjian, C.L. 1966. Candida albicans: saprophyte or pathogen? A diagnostic guide line. *J. Amer. Med. Ass.* 198:190-192.
25. Hurley, R. 1967. The pathogenic Candida species: A review. *Rev. Med. Vet. Mycol.* 6:159-176.
26. Kozinn, P.J.; Taschdjian, C.L. 1962. Enteric candidiasis: Diagnosis and clinical considerations. *Pediatrics* 30:71-85.
27. Emmons, C.W.; Binford, C.H.; Utz, J.P. 1964. Medical mycology, p. 131-144. Lea and Febiger, Philadelphia.
28. Winner, H.I.; Hurley, R. 1964. Candida albicans. Little, Brown and Co., Boston.

29. Vogel, R.A.; Koger, M.; Johnson, M.; Hunter, M. 1962. Tuberculin hypersensitivity associated with immunization of guinea pigs with Candida albicans and the presence of this organism in normal guinea pigs. Mycopathol. Mycol. Appl. 16:117-124.
30. Matsufuji, S.; Kukunda, M.; Nakamura, S. 1963. A common antigen between Candida albicans and Mycobacterium tuberculosis. Kurume Med. J. 10:225-231.

Unclassified

Security Classification

DOCUMENT CONTROL DATA - R & D		
(Security classification of title, body of abstract and indexing annotation must be entered when the overall report is classified)		
1. ORIGINATING ACTIVITY (Corporate author)		2a. REPORT SECURITY CLASSIFICATION
Department of the Army Fort Detrick, Frederick, Maryland, 21701		Unclassified
		2b. GROUP
3. REPORT TITLE		
CANDIDIASIS IN SIMIANS		
4. DESCRIPTIVE NOTES (Type of report and inclusive dates)		
5. AUTHOR(S) (First name, middle initial, last name)		
Steven E. Wikse Robert M. Kovatch James G. Fox		
6. REPORT DATE	7a. TOTAL NO. OF PAGES	7b. NO. OF REFS
September 1969	21	30
8a. CONTRACT OR GRANT NO.		8b. ORIGINATOR'S REPORT NUMBER(S)
a. PROJECT NO. 1B562602AD01		Technical Manuscript 559
c.		9b. OTHER REPORT NO(S) (Any other numbers that may be assigned this report)
d.		CMs 6592
10. DISTRIBUTION STATEMENT		
Qualified requesters may obtain copies of this publication from DDC. Foreign announcement and dissemination of this publication by DDC is not authorized. Release or announcement to the public is not authorized.		
11. SUPPLEMENTARY NOTES		12. SPONSORING MILITARY ACTIVITY
		Department of the Army Fort Detrick, Frederick, Maryland, 21701
13. ABSTRACT		
<p>Candidiasis was diagnosed in six monkeys over a 10-month period. Most cases had been on antibiotic therapy for enterocolitis. Fungal invasion was seen in epithelium of the tongue, oral cavity, esophagus, and colon, and in hard keratin of the nails. Gross lesions of the anterior alimentary tract were either white patches or ulcers of the mucosa. Lesions of the colon consisted of a thick pseudomembrane that contained numerous <u>Candida</u>. The nails exhibited typical <u>Candida onychomycosis</u>. <u>C. albicans</u> was isolated from the two cases that were cultured. Tissue invasion by <u>Candida</u> blastospores and hyphae was histologically demonstrated in all cases. (</p>		
14. Key Words		
<p>Candidiasis Monkeys Laboratory animals Fungus <u>Candida albicans</u> Histopathology</p>		

DD FORM 1473

REPLACES DD FORM 1473, 1 JAN 64, WHICH IS OBSOLETE FOR ARMY USE.

Unclassified
Security Classification